



# The interplay of effector and regulatory T cells in cancer

Rahul Roychoudhuri, Robert L Eil and Nicholas P Restifo

Regulatory T ( $T_{reg}$ ) cells suppress effector T ( $T_{eff}$ ) cells and prevent immune-mediated rejection of cancer. Much less appreciated are mechanisms by which  $T_{eff}$  cells antagonize  $T_{reg}$  cells. Herein, we consider how complex reciprocal interactions between  $T_{eff}$  and  $T_{reg}$  cells shape their population dynamics within tumors. Under states of tolerance, including during tumor escape, suppressed  $T_{eff}$  cells support  $T_{reg}$  cell populations through antigen-dependent provision of interleukin (IL)-2. During immune activation,  $T_{eff}$  cells can lose this supportive capacity and directly antagonize  $T_{reg}$  cell populations to neutralize their immunosuppressive function. While this latter state is rarely achieved spontaneously within tumors, we propose that therapeutic induction of immune activation has the potential to stably disrupt immunosuppressive population states resulting in durable cancer regression.

## Address

National Cancer Institute (NCI), National Institutes of Health, Bethesda, MD 20892, USA

Corresponding authors: Roychoudhuri, Rahul ([roychoudhuri@mail.nih.gov](mailto:roychoudhuri@mail.nih.gov)) and Restifo, Nicholas P ([restifo@nih.gov](mailto:restifo@nih.gov))

**Current Opinion in Immunology** 2015, **33**:101–111

This review comes from a themed issue on **Tumour immunology**

Edited by **Hans Schreiber** and **Philip D Greenberg**

For a complete overview see the [Issue](#) and the [Editorial](#)

Available online 27th February 2015

<http://dx.doi.org/10.1016/j.coi.2015.02.003>

0952-7915/Published by Elsevier Ltd.

## Introduction

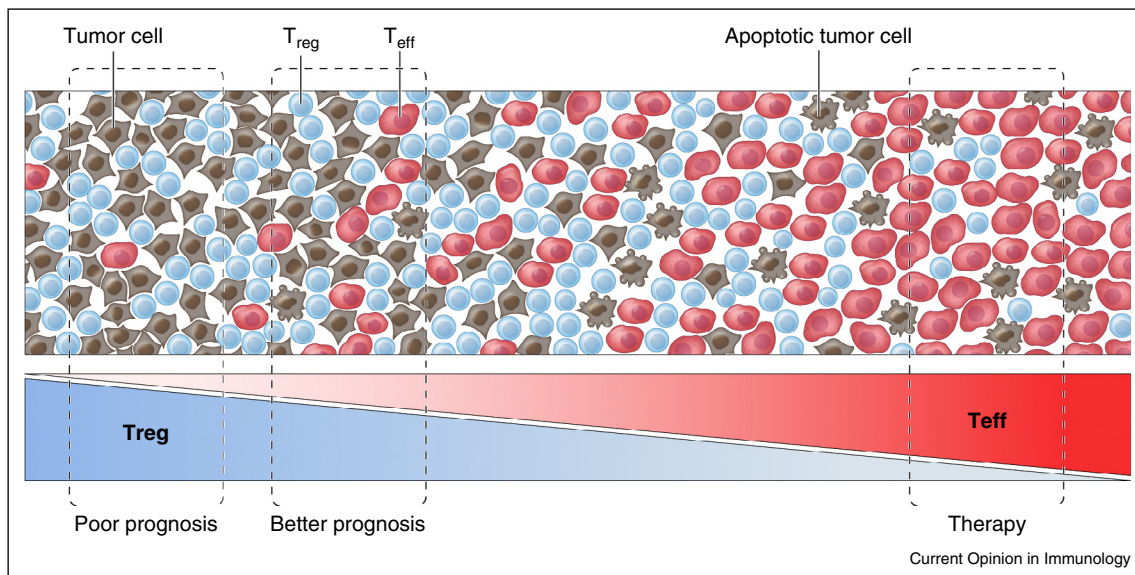
Growth of tumors in immunocompetent hosts is at odds with the powerful ability of the immune system to recognize and kill cancer cells. The cancer immunoediting hypothesis has been proposed as a conceptual framework to account for this behavior [1]. According to this hypothesis, tumor development is characterized by an initial ‘elimination’ phase, during which a majority of cancer cells are destroyed by various components of the immune system. This is followed by an ‘equilibrium’ phase, during which pressure from the immune system contributes to selection of tumor variants that give rise to an ‘escape’ phase characterized by evasion from immune control and unrestrained tumor growth. While selection of antigen-loss variants represents a mechanism of tumor escape and has been shown to contribute to growth of orthotopic

tumors [1–2], it fails to explain why established tumors continue to express immunogenic epitopes that are recognized by tumor-infiltrating lymphocytes and the efficacy of certain immune-based therapies for cancer [3–6,7<sup>\*\*</sup>,8]. Growth of tumors containing immunogenic epitopes is better explained through an understanding of the critical role of immunosuppression in promoting tumor escape [9–12]. Here, we review recent advances in our understanding of tumor immunosuppression and consider how a complex interplay between  $T_{reg}$  and  $T_{eff}$  cell populations dictates the outcome of tumor-specific immune responses.

A major advance in our understanding of peripheral tolerance arose with the identification of a suppressive subset of  $CD4^+$  T cells, referred to as  $T_{reg}$  cells, that express the high-affinity receptor for interleukin (IL)-2, IL-2R $\alpha$ , and whose deficiency in neonatally thymectomized mice results in lethal inflammatory disease [13]. The similar inflammatory phenotype manifested in ‘Scurfy’ mice [14] was later attributed to a complete defect in  $T_{reg}$  cell formation caused by an inactivating mutation within the gene encoding the transcription factor (TF) Forkhead box P3 (Foxp3). This resulted in identification of Foxp3 as a lineage specifying TF of  $T_{reg}$  cells [15–17]. A broader network of TFs, including BACH2 [18<sup>\*\*</sup>] and Foxo1 [19<sup>\*\*</sup>,20,21], are required establish the full  $T_{reg}$  cell transcriptional program. Humans lacking a functional *Foxp3* locus develop a lethal immune-mediated disease (immunodysregulation polyendocrinopathy enteropathy X-linked syndrome; IPEX) [22], while genetic polymorphisms within the *BACH2* locus are associated with multiple autoimmune and allergic diseases [18<sup>\*\*</sup>]. These findings provide evidence that  $T_{reg}$  cells regulate immune function in humans.

A complex interplay between  $CD4^+$   $T_{reg}$  and  $T_{eff}$  cells determines the outcome of immune reactions. Under homeostatic conditions,  $T_{reg}$  cells promote peripheral tolerance primarily through direct or indirect suppression of  $CD4^+$  Foxp3 $^-$   $T_{eff}$  cells. This is evidenced by the absence of inflammation in *Cd4*-deficient mice which lack both  $T_{reg}$  and  $CD4^+$   $T_{eff}$  cell populations [23].  $T_{reg}$  cells also suppress  $T_{eff}$  cell function within tumors (Figure 1). In murine tumor models, transient ablation of  $T_{reg}$  cells results in activation of  $CD4^+$  or  $CD8^+$   $T_{eff}$  cells and rejection of solid tumors [24,25,26<sup>\*\*</sup>,27]. In human tumors, low  $T_{reg}$  cell to  $T_{eff}$  cell ratios are associated with favorable survival in ovarian cancer [28,29], breast cancer [30], non-small cell lung carcinoma [31], hepatocellular carcinoma [32], renal cell carcinoma [33], pancreatic cancer [34], gastric cancer [35], cervical cancer [36]

Figure 1



The balance of  $T_{\text{eff}}$  and  $T_{\text{reg}}$  cells in tumors determines the functional outcome of immune responses.  $T_{\text{reg}}$  cells are represented in blue;  $T_{\text{eff}}$  cells are represented in red; tumor cells are depicted in brown. A spectrum of  $T_{\text{eff}}:T_{\text{reg}}$  cell ratios is depicted in the diagram going from left to right. Increased apoptosis is observed as the  $T_{\text{eff}}:T_{\text{reg}}$  cell ratio increases from left to right. The two boxes to the left represent the range of  $T_{\text{eff}}:T_{\text{reg}}$  cell ratios normally found within solid tumors. Within this range, the higher  $T_{\text{eff}}:T_{\text{reg}}$  cell ratios is associated with good prognosis in multiple cancer types (Refs. [23–25,26\*\*,27–32]). The desired outcome of therapeutic manipulation of  $T_{\text{eff}}:T_{\text{reg}}$  cell ratios is depicted in the box to the right, where predominant  $T_{\text{eff}}$  cell populations mediate widespread tumor cell destruction and eradication of disease.

and colorectal carcinoma [37].  $CD4^+$  and  $CD8^+$   $T_{\text{eff}}$  cells exert tumoricidal activity through multiple means that are reviewed extensively elsewhere [38,39]. Thus, the balance between  $T_{\text{reg}}$  and  $T_{\text{eff}}$  cells dictates the outcome of tumor-specific immune responses (Figure 1). It is therefore important to understand factors that affect the population dynamics of  $T_{\text{reg}}$  and  $T_{\text{eff}}$  cells within tumors.

### Population dynamics of $T_{\text{reg}}$ and $T_{\text{eff}}$ cells within tumors

The immunosuppressive function of  $T_{\text{reg}}$  cells is at odds with the capacity of the immune system to mediate brisk and functional  $T_{\text{eff}}$  cell responses against harmful pathogens in the context of acute infection. This implies that  $T_{\text{reg}}$  cell suppressive capacity may be neutralized during infection to promote clearance of disease. Emerging evidence presents multiple mechanisms by which  $T_{\text{eff}}$  cells antagonize the size and function of  $T_{\text{reg}}$  cell populations not only during infection, but also during inflammation and under specific conditions within tumors. Thus, while suppression of  $T_{\text{eff}}$  cells by a functionally predominant  $T_{\text{reg}}$  cell population represents the *status quo* during the 'escape' phase of tumor development, we propose that under specific conditions that either occur spontaneously or are induced therapeutically, a second state involving  $T_{\text{eff}}$ -mediated antagonism of  $T_{\text{reg}}$  cell populations can be induced that results in predominant  $T_{\text{eff}}$  cell function and clearance of disease.

### The role of IL-2 signaling in $T_{\text{reg}}$ and $T_{\text{eff}}$ cell population dynamics

IL-2 was originally characterized as a lymphocyte growth factor *in vitro* and was considered to have immunostimulatory function [40,41]. Subsequent characterization using gene-deficient mice led to the surprising finding that its non-redundant biological function *in vivo* is to restrain lethal inflammation [42,43]. This is attributable to a non-redundant requirement for IL-2 in  $T_{\text{reg}}$  cell survival [44–46]. IL-2-driven induction of the anti-apoptotic Bcl-2 family member Mcl-1 is critical for IL-2-dependent survival of  $T_{\text{reg}}$  cells [47\*\*].

### Suppressed $T_{\text{eff}}$ cells support $T_{\text{reg}}$ cells through paracrine IL-2 production

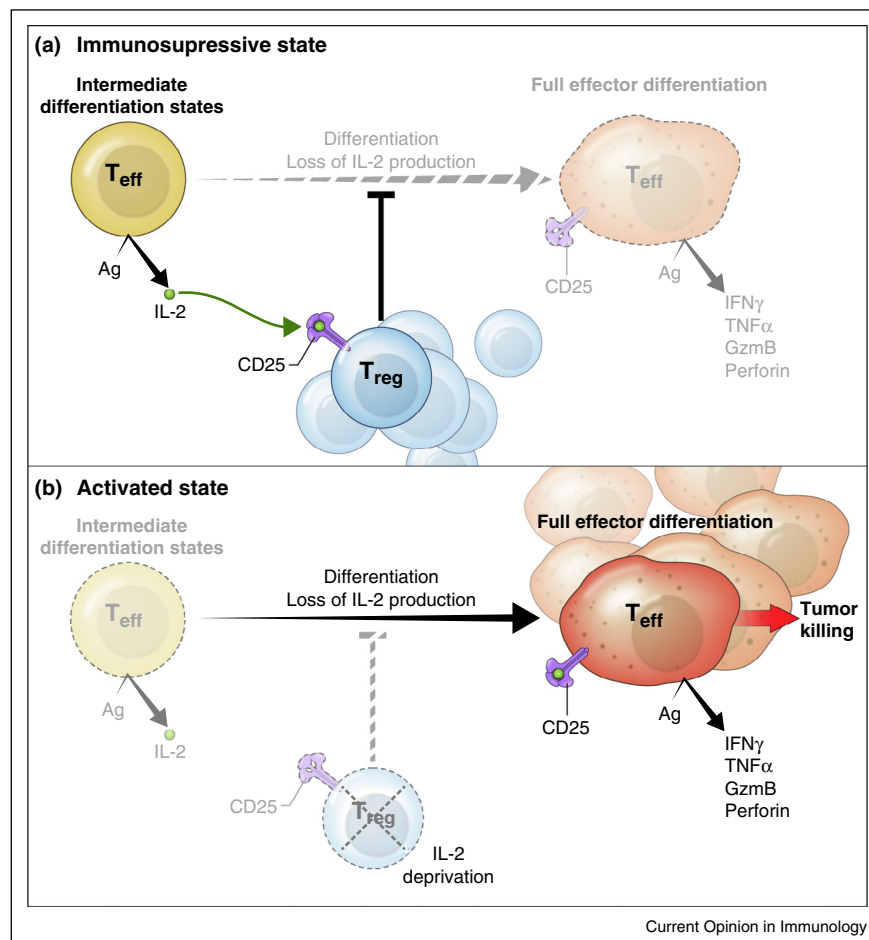
IL-2 is primarily produced by  $CD4^+$  T cells and  $CD8^+$  T cells and, to a lesser extent, NK cells. Since  $Foxp3^+$   $T_{\text{reg}}$  cells do not produce the cytokine, paracrine IL-2 generated in an antigen-dependent fashion by  $T_{\text{eff}}$  cells is required for  $T_{\text{reg}}$  cell survival [46,48]. This enables coupling of  $T_{\text{reg}}$  cell population size to the level of activation of  $T_{\text{eff}}$  cells at distinct sites providing a negative feedback loop that prevents excessive immune activation under homeostatic conditions [47\*\*,49,50\*\*]. However, not all  $T_{\text{eff}}$  cell populations produce IL-2 upon encounter with antigen and only ~10% of  $CD4^+$  T cells are primed for IL-2 production upon antigen stimulation [50\*\*]. In multiple T cell lineages, IL-2 expression is only associated

with activation of cells in an intermediate stage of differentiation and this is extinguished upon full effector differentiation. This process has been well characterized within the CD8<sup>+</sup> T cell lineage [51–54] and, to a lesser extent, within CD4<sup>+</sup> T cells [55,56]. Multiple signals, including IL-2 itself, contribute to loss of IL-2 production by T<sub>eff</sub> cell populations [50<sup>\*\*</sup>]. In part, this occurs through induction of the transcription factor PR domain zinc finger protein 1 (Blimp-1), that promotes acquisition of effector cell characteristics and loss of memory cell features including IL-2 production [56–60]. Thus, it is likely that T<sub>eff</sub> cells in intermediate stages of differentiation, rather than terminally differentiated effector cells, predominantly contribute to T<sub>reg</sub> cell maintenance [61<sup>\*\*</sup>].

It is possible that T<sub>reg</sub> cells actively maintain T<sub>eff</sub> cells in intermediate stages of differentiation by suppressing full

effector differentiation, resulting in stabilization of IL-2-producing populations of T<sub>eff</sub> cells. T<sub>reg</sub> cells play a critical role in formation of memory CD8<sup>+</sup> T cells by withholding IL-2 and preventing full effector differentiation in a subset of cells during the acute phase of primary CD8<sup>+</sup> T cell responses [62<sup>\*\*</sup>]. Moreover, production of TGF-β by T<sub>reg</sub> cells prevents full cytotoxic effector differentiation in tumor-specific CD8<sup>+</sup> T cells [63]. It is therefore plausible that T<sub>reg</sub> cells, through sequestration of IL-2 and production of TGF-β, suppress full effector differentiation in T<sub>eff</sub> cell populations to maintain a population of supportive IL-2-producing cells. This provides a potential feed-forward mechanism by which T<sub>reg</sub> cell populations are supported by suppressed T<sub>eff</sub> cells, constrained in intermediate stages of differentiation, to reinforce and stabilize the immunosuppressive state within tumors (Figure 2a).

Figure 2



IL-2: The currency of T<sub>reg</sub> and T<sub>eff</sub> cell population dynamics. **(a)** Immunosuppressive state. T<sub>reg</sub>-mediated suppression of T<sub>eff</sub> cells results in blockade of full effector differentiation. This maintains T<sub>eff</sub> cells in intermediate states of differentiation that support T<sub>reg</sub> cells through antigen-dependent paracrine IL-2 production. **(b)** Activated immune state. Reduced T<sub>reg</sub> cell suppression results in full T<sub>eff</sub> cell differentiation. This is accompanied by loss of antigen-dependent IL-2 production and withdrawal of IL-2 support for T<sub>reg</sub> cells. Additionally, expression of CD25 by activated T<sub>eff</sub> cells enables them to sequester any remaining IL-2.

### Unrestrained $T_{\text{eff}}$ cell differentiation results in withdrawal of paracrine IL-2 support

$T_{\text{eff}}$  cells progressively lose the capacity to produce IL-2 upon effector differentiation [50, 51–53, 55–60]. This raises the possibility that under certain conditions, unrestrained differentiation of  $T_{\text{eff}}$  cells results in withdrawal of cytokine support for  $T_{\text{reg}}$  cells. Such ‘withdrawal’ of cytokine support by  $T_{\text{eff}}$  cell populations is powerfully evidenced by the work of Oldenhove *et al.* in their study of  $T_{\text{reg}}$  and  $T_{\text{eff}}$  cell population dynamics during lethal infection with *Toxoplasma gondii* in mice [64]. Following infection, a striking collapse in  $T_{\text{reg}}$  cell frequency and absolute number coincides with loss of IL-2 production by  $\text{Foxp3}^{\text{+}} \text{CD4}^{\text{+}}$  T cells in the gut. This correlates with acquisition of a type helper 1 (Th1)-polarized state in which cells progressively lose expression of IL-2 and gain expression of IFN- $\gamma$  as infection progresses toward its lethal outcome. Strikingly, complementation of IL-2 signaling through provision of exogenous IL-2 restores  $T_{\text{reg}}$  cell numbers and prevents lethality. These findings have been recapitulated in *Listeria monocytogenes* and vaccinia virus infection [65]. Thus, induction of specific  $T_{\text{eff}}$  cell polarization states results in withdrawal of IL-2 support for  $T_{\text{reg}}$  cells, collapse in their population size, and unrestrained  $T_{\text{eff}}$  cell activation. Moreover, expression of IL-2R $\alpha$  on  $T_{\text{eff}}$  cells is induced by antigen activation and terminal differentiation and may enable  $T_{\text{eff}}$  cells to sequester any remaining IL-2, further limiting its availability to  $T_{\text{reg}}$  cells [66,67].

Thus, complex regulation of IL-2 and its high-affinity receptor IL-2R $\alpha$  results in the potential for bistability in  $T_{\text{reg}}$  and  $T_{\text{eff}}$  cell population dynamics within tumors. First, an immunosuppressive state exists where  $T_{\text{reg}}$  cells are supported by IL-2-producing  $T_{\text{eff}}$  cells constrained in intermediate states of differentiation (Figure 2a). This immunosuppressive state predominates during the escape phase of tumor development. Second, an activated immune state exists, in which unrestrained  $T_{\text{eff}}$  cell differentiation results in withdrawal of paracrine IL-2 support and competition for remaining IL-2 through antigen-driven expression of IL-2R $\alpha$  (Figure 2b). While this state is rarely achieved spontaneously following tumor escape, such conditions may be achieved following therapeutic intervention (discussed below). The potential for IL-2 competition to account for population bistability is supported by mathematical models [68–70].

### Reciprocal antagonism between $T_{\text{reg}}$ and $T_{\text{eff}}$ cell populations

#### $T_{\text{reg}}$ -mediated suppression of $T_{\text{eff}}$ cell populations

In states of tolerance, including during tumor escape,  $T_{\text{reg}}$  cells block the proliferation, survival and function of  $T_{\text{eff}}$  cells through multiple means. These have been extensively reviewed [71] and are depicted in Figure 3a. A subset of  $\text{Foxp3}^{\text{+}} \text{T}_{\text{reg}}$  cells constitutively express IL-2R $\alpha$  and sequestration of IL-2 by  $T_{\text{reg}}$  cells is a component of

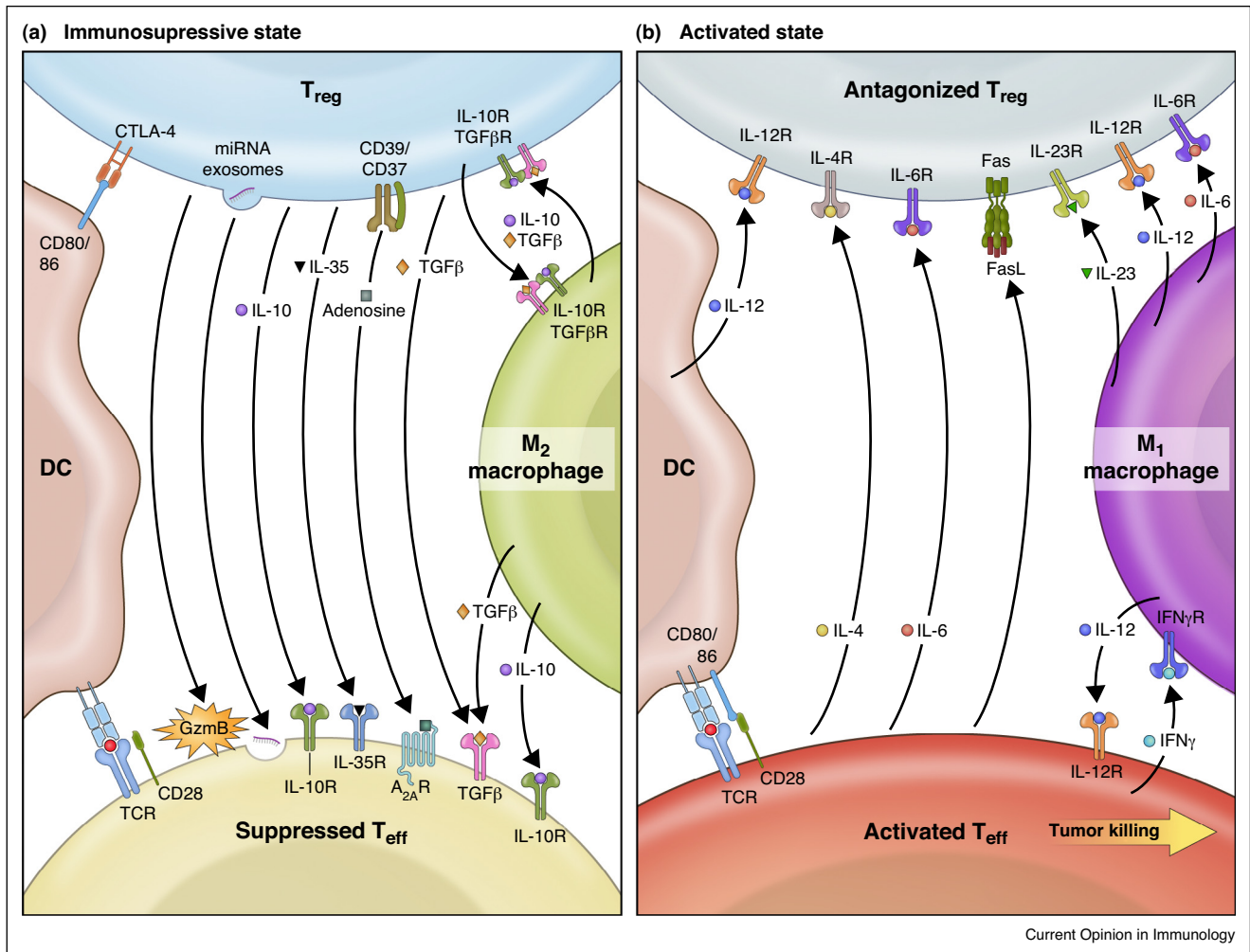
their suppressive function [72,73]. However, expansion of  $T_{\text{eff}}$  cells and induction of lethal inflammation caused by  $T_{\text{reg}}$  cell insufficiency can proceed in the complete absence of IL-2 since IL-2 deficient mice succumb to lethal autoimmunity related to defective  $T_{\text{reg}}$  cell homeostasis [42,46,66], indicating redundant modes of suppression. These have been extensively reviewed elsewhere [71] and include CTLA-4-mediated sequestration of CD80 and CD86 on the surface of APC by trogocytosis [74] and expression of inhibitory cytokines such as TGF- $\beta$  [75–77], IL-10 [78,79] and IL-35 [80,81]. Further, paracrine production of adenosine by  $T_{\text{reg}}$  cells suppresses  $T_{\text{eff}}$  cell differentiation and function [82,83]. Direct cytolysis of  $T_{\text{eff}}$  cells by  $T_{\text{reg}}$  cells is an additional mechanism of immunosuppression and has been attributed to expression of granzyme B and perforin by  $T_{\text{reg}}$  cells [84,85]. This was found to contribute to  $T_{\text{reg}}$ -mediated immunosuppression within tumors [86]. Finally, delivery of micro-RNA-containing exosomes is another mechanism of  $T_{\text{reg}}$ -mediated immunosuppression [87]. Innate immune cells contribute to feed-forward reinforcement of immunosuppressive states induced by  $T_{\text{reg}}$  cells. Amplification of inhibitory cytokine signaling by pro-tumorigenic tumor-associated macrophages (TAM) bearing an M2 (or alternatively activated) phenotype [88,89], and dendritic cells [9] are all implicated in promoting stability of immunosuppressive states within tumors.

#### Antagonism of $T_{\text{reg}}$ cell populations by $T_{\text{eff}}$ cells

During infection or upon induction of immune-mediated tumor rejection, emerging evidence indicates that  $T_{\text{eff}}$  cells may directly antagonize the stability and survival of  $T_{\text{reg}}$  cell populations to decrease their suppressive capacity, as depicted in Figure 3b. Selective killing of  $T_{\text{reg}}$  cells by  $T_{\text{eff}}$  cells is an example of such behavior. A decrease in  $T_{\text{reg}}$  cell numbers within tumors accompanies intratumoral IL-12 injection and this is dependent upon intrinsic expression of the death-receptor ligand FasL by  $\text{CD8}^{\text{+}}$  T cells [90]. This observation is attributed to direct FasL-mediated killing of  $T_{\text{reg}}$  cells by  $T_{\text{eff}}$  cells. Induction of  $T_{\text{reg}}$  cell lineage instability is another mechanism by which immune activation within tumors may antagonize  $T_{\text{reg}}$  cell population size. The topic of  $T_{\text{reg}}$  cell lineage instability is controversial. By indelibly labeling cells and their progeny that had transcriptionally activated the endogenous *Foxp3* locus, Rubtsov *et al.*, concluded that  $T_{\text{reg}}$  cells rarely convert into  $\text{Foxp3}^{\text{+}}$  ‘ex-Foxp3’ cells under physiological conditions or when perturbed during inflammation or lymphopenia [91]. However, using a similar approach based on a *Foxp3* reporter construct encoded by a bacterial artificial chromosome, Zhou *et al.*, observed accumulation of ‘ex-Foxp3’ IL-17-expressing cells in inflamed joints in response to synovial IL-6 [92]. Conversion of purified populations of  $\text{Foxp3}^{\text{+}} \text{T}_{\text{reg}}$  cells into  $\text{Foxp3}^{\text{+}}$  cells has also been observed in adoptive transfer models, both under conditions of extreme inflammation induced through allogeneic bone-marrow



Figure 3



Mechanisms of reciprocal antagonism between T<sub>reg</sub> and T<sub>eff</sub> cell populations. **(a)** Immunosuppressive state. T<sub>reg</sub> cells suppress T<sub>eff</sub> cell differentiation by multiple means including CTLA-4-dependent trogocytosis of CD80/86 on dendritic cells (DC), Granzyme B (GzmB) mediated killing of T<sub>eff</sub> cells, production of inhibitory cytokines and small molecules (adenosine, IL-10, TGF-β and miRNA exosomes). Additionally, macrophages and dendritic cells provide feed-forward reinforcement of immunosuppressive states. **(b)** Activated immune state. Activated T<sub>eff</sub> cells antagonize T<sub>reg</sub> cells under conditions of immune activation. T<sub>eff</sub> cells produce IL-6 and IFN-γ which leads to functional plasticity of T<sub>reg</sub> cells. T<sub>eff</sub> cell-derived IFN-γ causes dendritic cells and macrophages to produce IL-12 which results in a feed-forward cycle of IFN-γ production and induction of functional plasticity in T<sub>reg</sub> cells. Additionally, exposure to inflammatory cytokines may stimulate T<sub>reg</sub> cells to lose Foxp3 expression and acquire effector cell characteristics. Unrestrained differentiation and function of T<sub>eff</sub> cell populations leads to immune activation and tumor killing.

transplantation [93], and extensive lymphopenia-induced proliferation [94]. These studies implicate the inflammatory cytokines IL-6 and/or IL-4, that are produced by T<sub>eff</sub> cells, in driving lineage instability. It is therefore possible that under the inflammatory conditions induced by specific immune-based therapies for cancer, T<sub>reg</sub> cell lineage instability contributes to reversal of tumor immunosuppression.

The balance of T<sub>reg</sub> to T<sub>eff</sub> cells is also modulated at the level of their induction. A proportion of T<sub>reg</sub> cells found in peripheral tissues arise in the thymus (thymic T<sub>reg</sub> or

tT<sub>reg</sub> cells) and play a predominant role in promoting tumor immunosuppression [95]. However, induced T<sub>reg</sub> (iT<sub>reg</sub>) cells develop from conventional Foxp3<sup>-</sup> naive CD4<sup>+</sup> T cells in extrathymic tissues [96]. *De novo* induction of iT<sub>reg</sub> cells is observed within tumors [97] and their suppressive function has been demonstrated in the context of therapeutic vaccination [98] and adoptive transfer immunotherapy [99]. A number of effector cytokines, including IL-4, IL-6 and IL-23 are involved in driving *de novo* induction of T<sub>eff</sub> cell lineages from naive CD4<sup>+</sup> T cell precursors [100] and this process is implicitly reciprocal to *de novo* induction of iT<sub>reg</sub> cells from the same

precursor pool. Moreover, the  $T_{\text{eff}}$  cell cytokines IL-4, IFN- $\gamma$  and IL-23 directly antagonize TGF- $\beta$ -driven  $iT_{\text{reg}}$  cell induction [101–103].

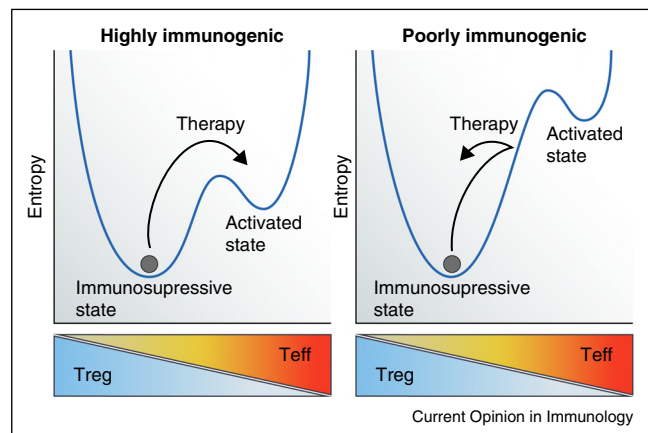
Finally, modulation of Foxp3<sup>+</sup>  $T_{\text{reg}}$  cell function is a distinct mechanism by which  $T_{\text{eff}}$  cells may alter the functional output of  $T_{\text{reg}}$  cell populations within tumors. Under conditions of lethal infection, Foxp3<sup>+</sup> cells expressing T-bet and IFN- $\gamma$  are induced as a result of high levels of IL-12 secretion by DC within the lamina propria [64]. Similar plasticity is observed during viral infection of the central nervous system in mice [104]. Inflammatory conditions can also directly antagonize the suppressive function of  $T_{\text{reg}}$  cells. Using a transfer colitis model, the inflammatory cytokine IL-23 was found to promote colitis by inhibiting the suppressive function of Foxp3<sup>+</sup>  $T_{\text{reg}}$  cells [103].

Thus, direct antagonism of  $T_{\text{reg}}$  cell number and function equip  $T_{\text{eff}}$  cells with the capacity to disrupt  $T_{\text{reg}}$ -mediated immunosuppression once a specific threshold of activation has been achieved. Feed-forward reinforcement by innate immune cells within tumors contributes to the stability of activated immune states. In particular, Th1 cytokine-induced polarization of TAM into M1 (or classically activated) cells further potentiates immune activation through production of IL-12, IL-1, IL-6, TNF- $\alpha$  and IL-23 [88,89]. Additionally, dendritic cells provide further potential for feed-forward reinforcement by producing IL-12 in response to  $T_{\text{eff}}$  cell-derived IFN- $\gamma$ .

### Opportunities for therapeutic intervention

Reciprocal antagonism and feed-forward reinforcement contribute to the potential for two distinct immune states. First, an immunosuppressive state, which is established early during the escape phase of tumor development, is stabilized through provision of IL-2 support by suppressed  $T_{\text{eff}}$  cells for  $T_{\text{reg}}$  cells. Second, an activated immune state, in which unrestrained  $T_{\text{eff}}$  cell differentiation is accompanied by withdrawal of cytokine support and direct antagonism of  $T_{\text{reg}}$  cell populations to drive clearance of disease. Despite the potential availability of activated immune states, immunosuppressive states represent the *status quo* of tumor escape and entry into activated immune states is rarely achieved. However, under certain circumstances that either arise spontaneously [105,106] or in the context of therapeutic manipulation, transition into a self-reinforcing activated immune state may occur. An entropy model can be utilized to consider the ‘energy-barrier’ to transition between immunosuppressive and activated immune states (Figure 4). The energy-barrier to transition between states may differ between highly immunogenic (left panel) and poorly immunogenic cancers (right panel). The molecular basis for differences between poorly immunogenic and highly immunogenic tumors are incompletely elucidated but new findings support a prominent contribution of

Figure 4



Bistable population dynamics of  $T_{\text{reg}}$  and  $T_{\text{eff}}$  cells within tumors. Entropy landscapes indicating bistable  $T_{\text{reg}}$  and  $T_{\text{eff}}$  cell population states in tumors. A stable low-entropy state involving functional predominance of  $T_{\text{reg}}$  cells over a minimal population of suppressed  $T_{\text{eff}}$  cells exists during the escape phase of tumor development (indicated by the position of the ball). Therapy (arrow) results in immune activation. *Left panel:* In the case of highly immunogenic tumors, an activation threshold is reached enabling stable transition into an activated immune state that results in tumor regression. *Right panel:* In the case of poorly immunogenic tumors, immune activation is insufficient to surmount the higher energy barrier to transition into an activated state resulting in stable persistence of immunosuppression and failure to clear disease.

mutational load in driving heterogeneous immunogenicity and clinical outcomes to immune-based therapy [107–109].

A bistable model of  $T_{\text{reg}}$  and  $T_{\text{eff}}$  cell population dynamics has two important implications for cancer immunotherapy. First, the potential for immune-based therapies to achieve durable complete regression of disease distinguishes them from traditional chemotherapy. An attractive explanation for the striking durability of such clinical responses despite transient administration of therapy is the induction of self-reinforcing activation states within tumors. Secondly, while direct targeting of  $T_{\text{reg}}$  cells represents a strategy for disruption of immunosuppression in cancer,  $T_{\text{eff}}$  cell populations directly and indirectly antagonize  $T_{\text{reg}}$  cells. Thus, induction of immune activation represents an alternate strategy to reverse immunosuppression and a potential mechanism by which some therapies targeting  $T_{\text{eff}}$  cell function cause durable regression of disease. Thus, current immune-based therapies must be considered in the context of their effect on  $T_{\text{reg}}$  and  $T_{\text{eff}}$  cell population dynamics within tumors.

Attempts have been made to distinguish the effect of immune-based therapies on  $T_{\text{reg}}$  and  $T_{\text{eff}}$  cell populations within tumors. However, a majority of current immune-based therapies directly affect both  $T_{\text{eff}}$  and  $T_{\text{reg}}$  cells.

Monoclonal antibody therapy targeting CTLA-4 results in durable complete responses in patients with metastatic melanoma [6]. Initial reports attributed efficacy to an isolated effect on  $T_{\text{eff}}$  cell populations [110–112]. However, CTLA-4 treatment results in increased  $T_{\text{eff}}:T_{\text{reg}}$  cell ratios within tumors [113<sup>•</sup>,114,115] and its efficacy, in part, requires depletion of antibody-bound  $T_{\text{reg}}$  cells by  $Fc\gamma$  receptor-expressing macrophages within the tumor microenvironment [116,117]. Thus, CTLA-4 therapy both activates  $T_{\text{eff}}$  cells and depletes  $T_{\text{reg}}$  cells within tumors and it is plausible that disruption of the immunosuppressive balance of  $T_{\text{reg}}$  and  $T_{\text{eff}}$  cells contributes to durable tumor regression observed in a subset of treated patients. Similarly, monoclonal antibody therapy targeting the programmed cell death 1 (PD-1) receptor results in durable regression of disease in a substantial proportion of patients with metastatic melanoma [7<sup>••</sup>] and in a smaller proportion of patients with renal cell carcinoma, non-small cell lung cancer and ovarian cancer [8,118]. PD-1 is preferentially expressed on recently activated or exhausted T cells and negatively regulates effector function [119]. However, PD-1 signaling is also implicated in the induction and function of peripheral  $iT_{\text{reg}}$  cells [120] and blockade of PD-1 signaling causes both decreased  $Foxp3^+ T_{\text{reg}}$  cell ratios and augmented  $T_{\text{eff}}$  cell function in a murine melanoma therapy model [121]. Thus, the striking efficacy of monoclonal antibodies targeting PD-1 and CTLA-4 may be related to alterations in the balance of  $T_{\text{reg}}$  and  $T_{\text{eff}}$  cells within tumors. IL-12 also causes profound immune-mediated tumor regression with concomitant activation of  $T_{\text{eff}}$  cells in preclinical models, though systemic delivery of IL-12 in humans results in limiting adverse events [122,123]. In a number of mouse models, IL-12 administration causes increased  $T_{\text{eff}}:T_{\text{reg}}$  cell ratios [124,125]. While in some models, this is attributable to killing of  $T_{\text{reg}}$  cells by  $T_{\text{eff}}$  cells within tumors [90], IL-12 also has direct effects on the function and stability of  $T_{\text{reg}}$  cells [64].

In some cases, the effects of immune-based therapies on  $T_{\text{reg}}:T_{\text{eff}}$  ratios are unclear. High-dose IL-2 therapy results in durable clearance of metastatic melanoma and renal cell carcinoma in a minority of patients [126–128]. While this therapy results in transiently increased frequencies of peripherally circulating  $T_{\text{reg}}$  cells [129], its long-term effect on the ratio of  $T_{\text{reg}}:T_{\text{eff}}$  cells following withdrawal of therapy and specifically within tumors is not well established. In mice, administration of exogenous IL-2 drives differentiation of  $T_{\text{eff}}$  cells and loss of the IL-2-producing population of cells [50<sup>••</sup>]. This is significant given the function of IL-2-producing  $T_{\text{eff}}$  cells in maintenance of  $T_{\text{reg}}$  cell populations. Depletion of the IL-2 producing  $T_{\text{eff}}$  cell pool within tumors and loss of  $T_{\text{reg}}$  cell supportive capacity amongst TIL may represent a mechanism by which high-dose IL-2 causes durable complete regression of disease in a subset of patients. Durable complete responses are also observed following

adoptive cell therapy (ACT) using bulk TIL populations [130–132] but the effect of therapy on endogenous  $T_{\text{reg}}$  and  $T_{\text{eff}}$  cell populations is poorly resolved. Reshaping of the endogenous  $T_{\text{reg}}:T_{\text{eff}}$  cell balance may, however, contribute to durable responses observed with both high-dose IL-2 therapy and ACT. Thus, a majority of immune-based therapies for cancer affect  $T_{\text{eff}}$  and  $T_{\text{reg}}$  cell population dynamics within tumors. Further detailed experimental investigation is required to separate direct from indirect effects of such therapies of  $T_{\text{reg}}$  and  $T_{\text{eff}}$  cell populations and their functional consequences.

In conclusion, mechanisms of reciprocal antagonism and self-reinforcement drive bistability in  $T_{\text{reg}}$  and  $T_{\text{eff}}$  cell population dynamics, enabling the immune system to exclusively consolidate the divergent outcomes of tolerance and immunity. An understanding of the complexity of  $T_{\text{reg}}$  and  $T_{\text{eff}}$  cell population dynamics has implications for rationalizing the striking durability and efficacy of immune-based therapies for cancer and provides a basis for development of new strategies that manipulate immune function in cancer patients.

## Acknowledgements and funding

The authors were supported by a generous gift from Li Jinyuan and the Tiens Charitable Foundation, the NIH-Center for Regenerative Medicine, the Milstein Family Foundation and by the Intramural Research Program of the National Cancer Institute (ZIA BC010763). R.R. is supported by a Sir Henry Dale Fellowship jointly funded by the Wellcome Trust and the Royal Society (Grant Number 105663/Z/14/Z). The authors thank Alan Hoofring and Ethan Tyler for their assistance with illustrations, and David Clever, Luca Gattinoni, Christopher Klebanoff, Yun Ji, Madhusudhanan Sukumar, Joseph Crompton and Jenny Pan for insightful comments and discussions.

## References and recommended reading

Papers of particular interest, published within the period of review, have been highlighted as:

- of special interest
- of outstanding interest

1. Dunn GP, Old LJ, Schreiber RD: **The three Es of cancer immunoediting.** *Annu Rev Immunol* 2004, **22**:329–360.
2. Dubey P, Hendrickson RC, Meredith SC, Siegel CT, Shabanowitz J, Skipper JC, Engelhard VH, Hunt DF, Schreiber H: **The immunodominant antigen of an ultraviolet-induced regressor tumor is generated by a somatic point mutation in the DEAD box helicase p68.** *J Exp Med* 1997, **185**:695–705.
3. Gros A, Robbins PF, Yao X, Li YF, Turcotte S, Tran E, Wunderlich JR, Mixon A, Farid S, Dudley ME *et al.*: **PD-1 identifies the patient-specific CD8(+) tumor-reactive repertoire infiltrating human tumors.** *J Clin Invest* 2014, **124**:2246–2259.
4. Tran E, Turcotte S, Gros A, Robbins PF, Lu YC, Dudley ME, Wunderlich JR, Somerville RP, Hogan K, Hinrichs CS *et al.*: **Cancer immunotherapy based on mutation-specific CD4<sup>+</sup> T cells in a patient with epithelial cancer.** *Science* 2014, **344**:641–645.
5. Dudley ME, Wunderlich JR, Robbins PF, Yang JC, Hwu P, Schwartzentruber DJ, Topalian SL, Sherry R, Restifo NP, Hübicki AM *et al.*: **Cancer regression and autoimmunity in patients after clonal repopulation with antitumor lymphocytes.** *Science* 2002, **298**:850–854.
6. Hodi FS, O'Day SJ, McDermott DF, Weber RW, Sosman JA, Haanen JB, Gonzalez R, Robert C, Schadendorf D, Hassel JC



*et al.*: **Improved survival with ipilimumab in patients with metastatic melanoma.** *N Engl J Med* 2010, **363**:711-723.

7. Hamid O, Robert C, Daud A, Hodi FS, Hwu WJ, Kefford R, Wolchok JD, Hersey P, Joseph RW, Weber JS *et al.*: **Safety and tumor responses with lambrolizumab (anti-PD-1) in melanoma.** *N Engl J Med* 2013, **369**:134-144.

This landmark study demonstrates efficacy of  $\alpha$ -PD-1 treatment in patients with metastatic melanoma.

8. Topalian SL, Hodi FS, Brahmer JR, Gettinger SN, Smith DC, McDermott DF, Powderly JD, Carvajal RD, Sosman JA, Atkins MB *et al.*: **Safety, activity, and immune correlates of anti-PD-1 antibody in cancer.** *N Engl J Med* 2012, **366**:2443-2454.
9. Rabinovich GA, Gabrilovich D, Sotomayor EM: **Immunosuppressive strategies that are mediated by tumor cells.** *Annu Rev Immunol* 2007, **25**:267-296.
10. Quezada SA, Peggs KS, Simpson TR, Allison JP: **Shifting the equilibrium in cancer immunoeediting: from tumor tolerance to eradication.** *Immunol Rev* 2011, **241**:104-118.
11. Gajewski TF, Schreiber H, Fu YX: **Innate and adaptive immune cells in the tumor microenvironment.** *Nat Immunol* 2013, **14**:1014-1022.
12. Klebanoff CA, Khong HT, Antony PA, Palmer DC, Restifo NP: **Sinks, suppressors and antigen presenters: how lymphodepletion enhances T cell-mediated tumor immunotherapy.** *Trends Immunol* 2005, **26**:111-117.
13. Sakaguchi S, Sakaguchi N, Asano M, Itoh M, Toda M: **Immunologic self-tolerance maintained by activated T cells expressing IL-2 receptor  $\alpha$ -chains (CD25) Breakdown of a single mechanism of self-tolerance causes various autoimmune diseases.** *J Immunol* 1995, **155**:1151-1164.
14. Godfrey VL, Wilkinson JE, Rinchik EM, Russell LB: **Fatal lymphoreticular disease in the scurfy (sf) mouse requires T cells that mature in a sf thymic environment: potential model for thymic education.** *Proc Natl Acad Sci U S A* 1991, **88**:5528-5532.
15. Hori S, Nomura T, Sakaguchi S: **Control of regulatory T cell development by the transcription factor Foxp3.** *Science* 2003, **299**:1057-1061.
16. Khattry R, Cox T, Yasayko SA, Ramsdell F: **An essential role for Scurfin in CD4+CD25+ T regulatory cells.** *Nat Immunol* 2003, **4**:337-342.
17. Fontenot JD, Gavin MA, Rudensky AY: **Foxp3 programs the development and function of CD4+CD25+ regulatory T cells.** *Nat Immunol* 2003, **4**:330-336.
18. Roychowdhuri R, Hirahara K, Mousavi K, Clever D, Klebanoff CA, Bonelli M, Sciume G, Zare H, Vahedi G, Dema B *et al.*: **BACH2 represses effector programs to stabilize T(reg)-mediated immune homeostasis.** *Nature* 2013, **498**:506-510.

The gene encoding the transcription factor BACH2 is a prominent susceptibility locus for multiple autoimmune and allergic diseases. This study established a non-redundant role for BACH2 in maintenance of immune homeostasis through its role in T<sub>reg</sub> cell development.

19. Ouyang W, Liao W, Luo CT, Yin N, Huse M, Kim MV, Peng M, Chan P, Ma Q, Mo Y *et al.*: **Novel Foxo1-dependent transcriptional programs control T(reg) cell function.** *Nature* 2012, **491**:554-559.

This study demonstrated that T<sub>reg</sub> cell-intrinsic expression of the transcription factor Foxo1 is required for T<sub>reg</sub>-mediated immune homeostasis.

20. Samstein RM, Arvey A, Josefowicz SZ, Peng X, Reynolds A, Sandstrom R, Neph S, Sabo P, Kim JM, Liao W *et al.*: **Foxp3 exploits a pre-existent enhancer landscape for regulatory T cell lineage specification.** *Cell* 2012, **151**:153-166.
21. Kerdlies YM, Stone EL, Beisner DR, McGargill MA, Ch'en IL, Stockmann C, Katayama CD, Hedrick SM: **Foxo transcription factors control regulatory T cell development and function.** *Immunity* 2010, **33**:890-904.
22. Bennett CL, Christie J, Ramsdell F, Brunkow ME, Ferguson PJ, Whitesell L, Kelly TE, Saulsbury FT, Chance PF, Ochs HD: **The immune dysregulation, polyendocrinopathy, enteropathy,**

**X-linked syndrome (IPEX) is caused by mutations of FOXP3.** *Nat Genet* 2001, **27**:20-21.

23. Rahemtulla A, Fung-Leung WP, Schilham MW, Kundig TM, Sambhara SR, Narendran A, Arabian A, Wakeham A, Paige CJ, Zinkernagel RM *et al.*: **Normal development and function of CD8+ cells but markedly decreased helper cell activity in mice lacking CD4.** *Nature* 1991, **353**:180-184.
24. Klages K, Mayer CT, Lahl K, Lodenkemper C, Teng MW, Ngiow SF, Smyth MJ, Hamann A, Huehn J, Sparwasser T: **Selective depletion of Foxp3+ regulatory T cells improves effective therapeutic vaccination against established melanoma.** *Cancer Res* 2010, **70**:7788-7799.
25. Teng MW, Swann JB, von Scheidt B, Sharkey J, Zerafa N, McLaughlin N, Yamaguchi T, Sakaguchi S, Darcy PK, Smyth MJ: **Multiple antitumor mechanisms downstream of prophylactic regulatory T-cell depletion.** *Cancer Res* 2010, **70**:2665-2674.
26. Bos PD, Plitas G, Rudra D, Lee SY, Rudensky AY: **Transient regulatory T cell ablation deters oncogene-driven breast cancer and enhances radiotherapy.** *J Exp Med* 2013, **210**:2435-2466.

Systemic depletion of T<sub>reg</sub> cells results in significantly attenuated growth and metastasis of established tumors. This is accompanied by increased tumor cell apoptosis and tumor clearance was dependent upon CD4<sup>+</sup>T<sub>eff</sub> cells. Tumor clearance was not further augmented by  $\alpha$ -CTLA-4 or  $\alpha$ -PD-1/PD-L1 treatment. This demonstrates a significant role for promotion of solid tumor growth by endogenous T<sub>reg</sub> cells.

27. Teng MW, Ngiow SF, von Scheidt B, McLaughlin N, Sparwasser T, Smyth MJ: **Conditional regulatory T-cell depletion releases adaptive immunity preventing carcinogenesis and suppressing established tumor growth.** *Cancer Res* 2010, **70**:7800-7809.
28. Sato E, Olson SH, Ahn J, Bundy B, Nishikawa H, Qian F, Jungbluth AA, Frosina D, Gnjatic S, Ambrosone C *et al.*: **Intraepithelial CD8+ tumor-infiltrating lymphocytes and a high CD8+/regulatory T cell ratio are associated with favorable prognosis in ovarian cancer.** *Proc Natl Acad Sci U S A* 2005, **102**:18538-18543.
29. Curiel TJ, Coukos G, Zou L, Alvarez X, Cheng P, Mottram P, Evdeemon-Hogan M, Conejo-Garcia JR, Zhang L, Burow M *et al.*: **Specific recruitment of regulatory T cells in ovarian carcinoma fosters immune privilege and predicts reduced survival.** *Nat Med* 2004, **10**:942-949.
30. Bates GJ, Fox SB, Han C, Leek RD, Garcia JF, Harris AL, Banham AH: **Quantification of regulatory T cells enables the identification of high-risk breast cancer patients and those at risk of late relapse.** *J Clin Oncol* 2006, **24**:5373-5380.
31. Petersen RP, Campa MJ, Sperlazza J, Conlon D, Joshi MB, Harpole DH Jr, Patz EF Jr: **Tumor infiltrating Foxp3+ regulatory T-cells are associated with recurrence in pathologic stage I NSCLC patients.** *Cancer* 2006, **107**:2866-2872.
32. Gao Q, Qiu SJ, Fan J, Zhou J, Wang XY, Xiao YS, Xu Y, Li YW, Tang ZY: **Intratumoral balance of regulatory and cytotoxic T cells is associated with prognosis of hepatocellular carcinoma after resection.** *J Clin Oncol* 2007, **25**:2586-2593.
33. Griffiths RW, Elkord E, Gilham DE, Ramani V, Clarke N, Stern PL, Hawkins RE: **Frequency of regulatory T cells in renal cell carcinoma patients and investigation of correlation with survival.** *Cancer Immunol Immunother* 2007, **56**:1743-1753.
34. Hiraoka N, Onozato K, Kosuge T, Hirohashi S: **Prevalence of FOXP3+ regulatory T cells increases during the progression of pancreatic ductal adenocarcinoma and its premalignant lesions.** *Clin Cancer Res* 2006, **12**:5423-5434.
35. Perrone G, Ruffini PA, Catalano V, Spino C, Santini D, Muretto P, Spoto C, Zingaretti C, Sisti V, Alessandrini P *et al.*: **Intratumoral FOXP3-positive regulatory T cells are associated with adverse prognosis in radically resected gastric cancer.** *Eur J Cancer* 2008, **44**:1875-1882.
36. Jordanova ES, Gorter A, Ayachi O, Prins F, Durrant LG, Kenter GG, van der Burg SH, Fleuren GJ: **Human leukocyte antigen class I MHC class I chain-related molecule A, and CD8+/regulatory T-cell ratio: which variable determines survival of cervical cancer patients?** *Clin Cancer Res* 2008, **14**:2028-2035.



37. Sinicrope FA, Rego RL, Ansell SM, Knutson KL, Foster NR, Sargent DJ: **Intraepithelial effector (CD3+)/regulatory (FoxP3+) T-cell ratio predicts a clinical outcome of human colon carcinoma.** *Gastroenterology* 2009, **137**:1270-1279.
38. Graubert TA, Ley TJ: **How do lymphocytes kill tumor cells?** *Clin Cancer Res* 1996, **2**:785-789.
39. Ikeda H, Old LJ, Schreiber RD: **The roles of IFN gamma in protection against tumor development and cancer immunoeediting.** *Cytokine Growth Factor Rev* 2002, **13**:95-109.
40. Morgan DA, Ruscetti FW, Gallo R: **Selective in vitro growth of T lymphocytes from normal human bone marrows.** *Science* 1976, **193**:1007-1008.
41. Gillis S, Smith KA: **Long term culture of tumour-specific cytotoxic T cells.** *Nature* 1977, **268**:154-156.
42. Sadlack B, Merz H, Schorle H, Schimpl A, Feller AC, Horak I: **Ulcerative colitis-like disease in mice with a disrupted interleukin-2 gene.** *Cell* 1993, **75**:253-261.
43. Sadlack B, Lohler J, Schorle H, Klebb G, Haber H, Sichel E, Noelle RJ, Horak I: **Generalized autoimmune disease in interleukin-2-deficient mice is triggered by an uncontrolled activation and proliferation of CD4<sup>+</sup> T cells.** *Eur J Immunol* 1995, **25**:3053-3059.
44. Malek TR, Yu A, Vincek V, Scibelli P, Kong L: **CD4 regulatory T cells prevent lethal autoimmunity in IL-2Rbeta-deficient mice. Implications for the nonredundant function of IL-2.** *Immunity* 2002, **17**:167-178.
45. Bayer AL, Yu A, Adeegbe D, Malek TR: **Essential role for interleukin-2 for CD4(+)CD25(+) T regulatory cell development during the neonatal period.** *J Exp Med* 2005, **201**:769-777.
46. Fontenot JD, Rasmussen JP, Gavin MA, Rudensky AY: **A function for interleukin 2 in Foxp3-expressing regulatory T cells.** *Nat Immunol* 2005, **6**:1142-1151.
47. Pierson W, Cauwe B, Policheni A, Schlenner SM, Franckaert D, Berges J, Humblet-Baron S, Schonfeldt S, Herold MJ, Hildeman D *et al.*: **Antiapoptotic Mcl-1 is critical for the survival and niche-filling capacity of Foxp3(+) regulatory T cells.** *Nat Immunol* 2013, **14**:959-965.
- Incomplete depletion of endogenous T<sub>reg</sub> cells is accompanied by expansion of the residual T<sub>reg</sub> cell pool. This is driven by increased production of IL-2 from conventional T cells. IL-2 acts to upregulate Mcl-1 within T<sub>reg</sub> cells and conditional deletion of Mcl-1 results in defective T<sub>reg</sub> cell homeostasis and fatal autoimmunity.
48. Yu A, Zhu L, Altman NH, Malek TR: **A low interleukin-2 receptor signaling threshold supports the development and homeostasis of T regulatory cells.** *Immunity* 2009, **30**:204-217.
49. Almeida AR, Zaragoza B, Freitas AA: **Indexation as a novel mechanism of lymphocyte homeostasis: the number of CD4+CD25+ regulatory T cells is indexed to the number of IL-2-producing cells.** *J Immunol* 2006, **177**:192-200.
50. Amado IF, Berges J, Luther RJ, Mailhe MP, Garcia S, Bandeira A, Weaver C, Liston A, Freitas AA: **IL-2 coordinates IL-2-producing and regulatory T cell interplay.** *J Exp Med* 2013, **210**:2707-2720.
- The compartment of IL-2 producing CD4<sup>+</sup> T cells is tightly controlled and is comprises approximately 10% of total CD4<sup>+</sup> T cells under homeostatic conditions in mice. Administration of exogenous IL-2 decreases the size of the IL-2-producing population while depletion of T<sub>reg</sub> cells increases the number of IL-2-producing cells. This provides a negative feedback loop to regulate the size of the T<sub>reg</sub> cell pool and enable its coupling to the activation state of T<sub>eff</sub> cells under physiological conditions.
51. Wherry EJ, Teichgraber V, Becker TC, Masopust D, Kaech SM, Antia R, von Andrian UH, Ahmed R: **Lineage relationship and protective immunity of memory CD8 T cell subsets.** *Nat Immunol* 2003, **4**:225-234.
52. Buchholz VR, Flossdorf M, Hensel I, Kretschmer L, Weissbrich B, Graf P, Verschoor A, Schiemann M, Hofer T, Busch DH: **Disparate individual fates compose robust CD8+ T cell immunity.** *Science* 2013, **340**:630-635.
53. Klebanoff CA, Gattinoni L, Restifo NP: **CD8+ T-cell memory in tumor immunology and immunotherapy.** *Immunol Rev* 2006, **211**:214-224.
54. Roychoudhuri R, Lefebvre F, Honda M, Pan L, Ji Y, Klebanoff CA, Nichols CN, Fourati S, Hegazy AN, Goulet JP *et al.*: **Transcriptional profiles reveal a stepwise developmental program of memory CD8 T cell differentiation.** *Vaccine* 2014.
55. Seder RA, Darrah PA, Roederer M: **T-cell quality in memory and protection: implications for vaccine design.** *Nat Rev Immunol* 2008, **8**:247-258.
56. Pantaleo G, Harari A: **Functional signatures in antiviral T-cell immunity for monitoring virus-associated diseases.** *Nat Rev Immunol* 2006, **6**:417-423.
57. Shin H, Blackburn SD, Intlekofer AM, Kao C, Angelosanto JM, Reiner SL, Wherry EJ: **A role for the transcriptional repressor Blimp-1 in CD8(+) T cell exhaustion during chronic viral infection.** *Immunity* 2009, **31**:309-320.
58. Rutishauser RL, Martins GA, Kalachikov S, Chandele A, Parish IA, Meffre E, Jacob J, Calame K, Kaech SM: **Transcriptional repressor Blimp-1 promotes CD8(+) T cell terminal differentiation and represses the acquisition of central memory T cell properties.** *Immunity* 2009, **31**:296-308.
59. Xin A, Nutt SL, Belz GT, Kallies A: **Blimp1: driving terminal differentiation to a T.** *Adv Exp Med Biol* 2011, **780**:85-100.
60. Gong D, Malek TR: **Cytokine-dependent Blimp-1 expression in activated T cells inhibits IL-2 production.** *J Immunol* 2007, **178**:242-252.
61. Liston A, Gray DH: **Homeostatic control of regulatory T cell diversity.** *Nat Rev Immunol* 2014, **14**:154-165.
- In this important review, the role of IL-2 in driving distinctive T<sub>reg</sub> and T<sub>eff</sub> cell population dynamics under various physiological and disease states is discussed.
62. de Goer de Herve MG, Jaafoura S, Vallee M, Taoufik Y: **FoxP3(+) regulatory CD4 T cells control the generation of functional CD8 memory.** *Nat Commun* 2012, **3**:986.
- In this study, depletion of T<sub>reg</sub> cells during the acute phase of vaccinia virus infection results in decreased ability to mount secondary recall responses upon reinfection. The presence of T<sub>reg</sub> cells decreases exposure of CD8+ T cells to IL-2 during priming enabling generation of long-lived memory cell responses. This is significant to the topic of this review since memory CD8+ T cells produce higher levels of IL-2 upon antigen-activation than effector cells and defines a function of T<sub>reg</sub> cells in their differentiation.
63. Chen ML, Pittet MJ, Gorelik L, Flavell RA, Weissleder R, von Boehmer H, Khazaie K: **Regulatory T cells suppress tumor-specific CD8 T cell cytotoxicity through TGF-beta signals in vivo.** *Proc Natl Acad Sci U S A* 2005, **102**:419-424.
64. Oldenhove G, Bouladoux N, Wohlfert EA, Hall JA, Chou D, Dos Santos L, O'Brien S, Blank R, Lamb E, Natarajan S *et al.*: **Decrease of Foxp3<sup>+</sup> T<sub>reg</sub> cell number and acquisition of effector cell phenotype during lethal infection.** *Immunity* 2009, **31**:772-786.
65. Benson A, Murray S, Divakar P, Burnaevskiy N, Pifer R, Forman J, Yarovinsky F: **Microbial infection-induced expansion of effector T cells overcomes the suppressive effects of regulatory T cells via an IL-2 deprivation mechanism.** *J Immunol* 2012, **188**:800-810.
- This study extends earlier work by Oldenhove *et al.* [64] and provides evidence that during parasitic, bacterial, and viral infection T<sub>eff</sub> cells differentiate into a highly activated state in which they produce IFN- $\gamma$ , but lose the capacity to produce IL-2. Loss of IL-2 availability results in reduced T<sub>reg</sub> cell population size and suppressive capacity and this is reversed by supplementation with exogenous IL-2.
66. Malek TR, Castro I: **Interleukin-2 receptor signaling: at the interface between tolerance and immunity.** *Immunity* 2010, **33**:153-165.
67. Kalia V, Sarkar S, Subramaniam S, Haining WN, Smith KA, Ahmed R: **Prolonged interleukin-2Ralpha expression on virus-specific CD8+ T cells favors terminal-effector differentiation in vivo.** *Immunity* 2010, **32**:91-103.
68. Busse D, de la Rosa M, Hobiger K, Thurley K, Flossdorf M, Scheffold A, Hofer T: **Competing feedback loops shape IL-2 signaling between helper and regulatory T lymphocytes in cellular microenvironments.** *Proc Natl Acad Sci U S A* 2010, **107**:3058-3063.

69. Feinerman O, Jentsch G, Tkach KE, Coward JW, Hathorn MM, Sneddon MW, Emonet T, Smith KA, Altan-Bonnet G: **Single-cell quantification of IL-2 response by effector and regulatory T cells reveals critical plasticity in immune response.** *Mol Syst Biol* 2010, **6**:437.
70. Almeida AR, Amado IF, Reynolds J, Berges J, Lythe G, Molinaro Paris C, Freitas AA: **Quorum-sensing in CD4(+) T cell homeostasis: a hypothesis and a model.** *Front Immunol* 2012, **3**:125.
71. Vignali DA, Collison LW, Workman CJ: **How regulatory T cells work.** *Nat Rev Immunol* 2008, **8**:523-532.
72. Thornton AM, Shevach EM: **CD4+CD25+ immunoregulatory T cells suppress polyclonal T cell activation in vitro by inhibiting interleukin 2 production.** *J Exp Med* 1998, **188**:287-296.
73. McNally A, Hill GR, Sparwasser T, Thomas R, Steptoe RJ: **CD4+CD25+ regulatory T cells control CD8+ T-cell effector differentiation by modulating IL-2 homeostasis.** *Proc Natl Acad Sci U S A* 2011, **108**:7529-7534.
74. Wing K, Onishi Y, Prieto-Martin P, Yamaguchi T, Miyara M, Fehervari Z, Nomura T, Sakaguchi S: **CTLA-4 control over Foxp3<sup>+</sup> regulatory T cell function.** *Science* 2008, **322**:271-275.
75. Nakamura K, Kitani A, Strober W: **Cell contact-dependent immunosuppression by CD4(+)/CD25(+) regulatory T cells is mediated by cell surface-bound transforming growth factor beta.** *J Exp Med* 2001, **194**:629-644.
76. Marie JC, Liggitt D, Rudensky AY: **Cellular mechanisms of fatal early-onset autoimmunity in mice with the T cell-specific targeting of transforming growth factor-beta receptor.** *Immunity* 2006, **25**:441-454.
77. Gorelik L, Flavell RA: **Immune-mediated eradication of tumors through the blockade of transforming growth factor-beta signaling in T cells.** *Nat Med* 2001, **7**:1118-1122.
78. Asseman C, Mauze S, Leach MW, Coffman RL, Powrie F: **An essential role for interleukin 10 in the function of regulatory T cells that inhibit intestinal inflammation.** *J Exp Med* 1999, **190**:995-1004.
79. Chaudhry A, Samstein RM, Treuting P, Liang Y, Pils MC, Heinrich JM, Jack RS, Wunderlich FT, Bruning JC, Muller W *et al.*: **Interleukin-10 signaling in regulatory T cells is required for suppression of Th17 cell-mediated inflammation.** *Immunity* 2011, **34**:566-578.
80. Collison LW, Workman CJ, Kuo TT, Boyd K, Wang Y, Vignali KM, Cross R, Sehy D, Blumberg RS, Vignali DA: **The inhibitory cytokine IL-35 contributes to regulatory T-cell function.** *Nature* 2007, **450**:566-569.
81. Collison LW, Delgoffe GM, Guy CS, Vignali KM, Chaturvedi V, Fairweather D, Satoskar AR, Garcia KC, Hunter CA, Drake CG *et al.*: **The composition and signaling of the IL-35 receptor are unconventional.** *Nat Immunol* 2012, **13**:290-299.
82. Deaglio S, Dwyer KM, Gao W, Friedman D, Usheva A, Erat A, Chen JF, Enjoji K, Linden J, Oukka M *et al.*: **Adenosine generation catalyzed by CD39 and CD73 expressed on regulatory T cells mediates immune suppression.** *J Exp Med* 2007, **204**:1257-1265.
83. Kobie JJ, Shah PR, Yang L, Rebhahn JA, Fowell DJ, Mosmann TR: **T regulatory and primed uncommitted CD4 T cells express CD73, which suppresses effector CD4 T cells by converting 5'-adenosine monophosphate to adenosine.** *J Immunol* 2006, **177**:6780-6786.
84. Grossman WJ, Verbsky JW, Barchet W, Colonna M, Atkinson JP, Ley TJ: **Human T regulatory cells can use the perforin pathway to cause autologous target cell death.** *Immunity* 2004, **21**:589-601.
85. Gondek DC, Lu LF, Quezada SA, Sakaguchi S, Noelle RJ: **Cutting edge: contact-mediated suppression by CD4+CD25+ regulatory cells involves a granzyme B-dependent, perforin-independent mechanism.** *J Immunol* 2005, **174**:1783-1786.
86. Cao X, Cai SF, Fehniger TA, Song J, Collins LI, Piwnicka-Worms DR, Ley TJ: **Granzyme B and perforin are important for regulatory T cell-mediated suppression of tumor clearance.** *Immunity* 2007, **27**:635-646.
87. Okoye IS, Coomes SM, Pelly VS, Czieso S, Papayannopoulos V, Tolmachova T, Seabra MC, Wilson MS: **MicroRNA-containing T-regulatory-cell-derived exosomes suppress pathogenic T helper 1 cells.** *Immunity* 2014, **41**:89-103.
88. Noy R, Pollard JW: **Tumor-associated macrophages: from mechanisms to therapy.** *Immunity* 2014, **41**:49-61.
89. Biswas SK, Mantovani A: **Macrophage plasticity and interaction with lymphocyte subsets: cancer as a paradigm.** *Nat Immunol* 2010, **11**:889-896.
90. Kilinc MO, Rowsewell-Turner RB, Gu T, Virtuoso LP, Egilmez NK: **Activated CD8+ T-effector/memory cells eliminate CD4+CD25+ Foxp3+ T-suppressor cells from tumors via FasL mediated apoptosis.** *J Immunol* 2009, **183**:7656-7660.
91. Rubtsov YP, Nieuwehuis RE, Josefowicz S, Li L, Darce J, Mathis D, Benoist C, Rudensky AY: **Stability of the regulatory T cell lineage in vivo.** *Science* 2010, **329**:1667-1671.
92. Zhou X, Bailey-Bucktrout SL, Jeker LT, Penaranda C, Martinez-Llordella M, Ashby M, Nakayama M, Rosenthal W, Bluestone JA: **Instability of the transcription factor Foxp3 leads to the generation of pathogenic memory T cells in vivo.** *Nat Immunol* 2009, **10**:1000-1007.
93. Laurence A, Amarnath S, Mariotti J, Kim YC, Foley J, Eckhaus M, O'Shea JJ, Fowler DH: **STAT3 transcription factor promotes instability of nT<sub>reg</sub> cells and limits generation of iT<sub>reg</sub> cells during acute murine graft-versus-host disease.** *Immunity* 2012, **37**:209-222.
94. Komatsu N, Mariotti-Ferrandiz ME, Wang Y, Malissen B, Waldmann H, Hori S: **Heterogeneity of natural Foxp3<sup>+</sup> T cells: a committed regulatory T-cell lineage and an uncommitted minor population retaining plasticity.** *Proc Natl Acad Sci U S A* 2009, **106**:1903-1908.
95. Malchow S, Leventhal DS, Nishi S, Fischer BI, Shen L, Paner GP, Amit AS, Kang C, Geddes JE, Allison JP *et al.*: **Aire-dependent thymic development of tumor-associated regulatory T cells.** *Science* 2013, **339**:1219-1224.
96. Josefowicz SZ, Nieuwehuis RE, Kim HY, Treuting P, Chinen T, Zheng Y, Umetsu DT, Rudensky AY: **Extrathymically generated regulatory T cells control mucosal Th2 inflammation.** *Nature* 2012, **482**:395-399.
97. Zhou G, Levitsky HI: **Natural regulatory T cells and de novo-induced regulatory T cells contribute independently to tumor-specific tolerance.** *J Immunol* 2007, **178**:2155-2162.
98. Schreiber TH, Wolf D, Bodero M, Podack E: **Tumor antigen specific iT<sub>reg</sub> accumulate in the tumor microenvironment and suppress therapeutic vaccination.** *Oncimmunology* 2012, **1**:642-648.
99. Goding SR, Wilson KA, Xie Y, Harris KM, Baxi A, Akpinarli A, Fulton A, Tamada K, Strome SE, Antony PA: **Restoring immune function of tumor-specific CD4<sup>+</sup> T cells during recurrence of melanoma.** *J Immunol* 2013, **190**:4899-4909.
100. O'Shea JJ, Paul WE: **Mechanisms underlying lineage commitment and plasticity of helper CD4<sup>+</sup> T cells.** *Science* 2010, **327**:1098-1102.
101. Wei J, Duramad O, Perng OA, Reiner SL, Liu YJ, Qin FX: **Antagonistic nature of T helper 1/2 developmental programs in opposing peripheral induction of Foxp3<sup>+</sup> regulatory T cells.** *Proc Natl Acad Sci U S A* 2007, **104**:18169-18177.
102. Mantel PY, Kuipers H, Boyman O, Rhyner C, Ouaked N, Ruckert B, Karagiannidis C, Lambrecht BN, Hendriks RW, Cramer R *et al.*: **GATA3-driven Th2 responses inhibit TGF-beta1-induced FOXP3 expression and the formation of regulatory T cells.** *PLoS Biol* 2007, **5**:e329.
103. Izcue A, Hue S, Buonocore S, Arancibia-Carcamo CV, Ahern PP, Iwakura Y, Maloy KJ, Powrie F: **Interleukin-23 restrains regulatory T cell activity to drive T cell-dependent colitis.** *Immunity* 2008, **28**:559-570.

104. Zhao J, Fett C, Trandem K, Fleming E, Perlman S: **IFN-gamma-IL-10-expressing virus epitope-specific Foxp3(+) T reg cells in the central nervous system during encephalomyelitis.** *J Exp Med* 2011, **208**:1571-1577.
105. Kalialis LV, Drzewiecki KT, Mohammadi M, Mehlsen AB, Klyver H: **Spontaneous regression of metastases from malignant melanoma: a case report.** *Melanoma Res* 2008, **18**:279-283.
106. Bramhall RJ, Mahady K, Peach AH: **Spontaneous regression of metastatic melanoma — clinical evidence of the abscopal effect.** *Eur J Surg Oncol* 2014, **40**:34-41.
107. Alexandrov LB, Nik-Zainal S, Wedge DC, Aparicio SA, Behjati S, Biankin AV, Bignell GR, Bolli N, Borg A, Borresen-Dale AL *et al.*: **Signatures of mutational processes in human cancer.** *Nature* 2013, **500**:415-421.
108. Gubin MM, Zhang X, Schuster H, Caron E, Ward JP, Nogueira T, Ivanova Y, Hundal J, Arthur CD, Krebber WJ *et al.*: **Checkpoint blockade cancer immunotherapy targets tumour-specific mutant antigens.** *Nature* 2014, **515**:577-581.
109. Snyder A, Makarov V, Merghoub T, Yuan J, Zaretsky JM, Desrichard A, Walsh LA, Postow MA, Wong P, Ho TS *et al.*: **Genetic basis for clinical response to CTLA-4 blockade in melanoma.** *N Engl J Med* 2014, **371**:2189-2199.
110. Leach DR, Krummel MF, Allison JP: **Enhancement of antitumor immunity by CTLA-4 blockade.** *Science* 1996, **271**:1734-1736.
111. Chambers CA, Krummel MF, Boitell B, Hurwitz A, Sullivan TJ, Fournier S, Cassell D, Brunner M, Allison JP: **The role of CTLA-4 in the regulation and initiation of T-cell responses.** *Immunol Rev* 1996, **153**:27-46.
112. Suttmüller RP, van Duivenvoorde LM, van Elsas A, Schumacher TN, Wildenberg ME, Allison JP, Toes RE, Offringa R, Melief CJ: **Synergism of cytotoxic T lymphocyte-associated antigen 4 blockade and depletion of CD25(+) regulatory T cells in antitumor therapy reveals alternative pathways for suppression of autoreactive cytotoxic T lymphocyte responses.** *J Exp Med* 2001, **194**:823-832.
113. Quezada SA, Peggs KS, Curran MA, Allison JP: **CTLA4 blockade and GM-CSF combination immunotherapy alters the intratumor balance of effector and regulatory T cells.** *J Clin Invest* 2006, **116**:1935-1945.
- Blockade of CTLA-4 function in T<sub>eff</sub> cells is considered to be a mode of anti-tumor activity of  $\alpha$ -CTLA-4 treatment. In this study, the authors elegantly demonstrate that  $\alpha$ -CTLA-4 treatment functions, in part, by causing antibody-mediated depletion of intratumoral T<sub>reg</sub> cells by Fc $\gamma$ R-IV-expressing macrophages. This implies that CTLA-4 therapy has distinct effects of T<sub>eff</sub> and T<sub>reg</sub> cell populations in tumors.
114. Liakou CI, Kamat A, Tang DN, Chen H, Sun J, Troncoso P, Logothetis C, Sharma P: **CTLA-4 blockade increases IFN-gamma-producing CD4+ICOShi cells to shift the ratio of effector to regulatory T cells in cancer patients.** *Proc Natl Acad Sci U S A* 2008, **105**:14987-14999.
115. Waitz R, Solomon SB, Petre EN, Trumble AE, Fasso M, Norton L, Allison JP: **Potent induction of tumor immunity by combining tumor cryoablation with anti-CTLA-4 therapy.** *Cancer Res* 2012, **72**:430-439.
116. Simpson TR, Li F, Montalvo-Ortiz W, Sepulveda MA, Bergerhoff K, Arce F, Roddie C, Henry JY, Yagita H, Wolchok JD *et al.*: **Fc-dependent depletion of tumor-infiltrating regulatory T cells co-defines the efficacy of anti-CTLA-4 therapy against melanoma.** *J Exp Med* 2013, **210**:1695-1710.
117. Peggs KS, Quezada SA, Chambers CA, Korman AJ, Allison JP: **Blockade of CTLA-4 on both effector and regulatory T cell compartments contributes to the antitumor activity of anti-CTLA-4 antibodies.** *J Exp Med* 2009, **206**:1717-1725.
118. Brahmer JR, Tykodi SS, Chow LQ, Hwu WJ, Topalian SL, Hwu P, Drake CG, Camacho LH, Kauh J, Odunsi K *et al.*: **Safety and activity of anti-PD-L1 antibody in patients with advanced cancer.** *N Engl J Med* 2012, **366**:2455-2465.
119. Barber DL, Wherry EJ, Masopust D, Zhu B, Allison JP, Sharpe AH, Freeman GJ, Ahmed R: **Restoring function in exhausted CD8 T cells during chronic viral infection.** *Nature* 2006, **439**:682-687.
120. Francisco LM, Salinas VH, Brown KE, Vanguri VK, Freeman GJ, Kuchroo VK, Sharpe AH: **PD-L1 regulates the development, maintenance, and function of induced regulatory T cells.** *J Exp Med* 2009, **206**:3015-3029.
121. Zhou Q, Xiao H, Liu Y, Peng Y, Hong Y, Yagita H, Chandler P, Munn DH, Mellor A, Fu N *et al.*: **Blockade of programmed death-1 pathway rescues the effector function of tumor-infiltrating T cells and enhances the antitumor efficacy of lentivector immunization.** *J Immunol* 2010, **185**:5082-5092.
122. Colombo MP, Trinchieri G: **Interleukin-12 in anti-tumor immunity and immunotherapy.** *Cytokine Growth Factor Rev* 2002, **13**:155-168.
123. Chinnasamy D, Yu Z, Kerkar SP, Zhang L, Morgan RA, Restifo NP, Rosenberg SA: **Local delivery of interleukin-12 using T cells targeting VEGF receptor-2 eradicates multiple vascularized tumors in mice.** *Clin Cancer Res* 2012, **18**:1672-1683.
124. Vom Berg J, Vrohings M, Haller S, Haimovici A, Kulig P, Sledzinska A, Weller M, Becher B: **Intratumoral IL-12 combined with CTLA-4 blockade elicits T cell-mediated glioma rejection.** *J Exp Med* 2013, **210**:2803-2811.
125. Kilinc MO, Aulakh KS, Nair RE, Jones SA, Alard P, Kosiewicz MM, Egilmez NK: **Reversing tumor immune suppression with intratumoral IL-12: activation of tumor-associated T effector/memory cells, induction of T suppressor apoptosis, and infiltration of CD8+ T effectors.** *J Immunol* 2006, **177**:6962-6973.
126. Yang JC, Topalian SL, Parkinson D, Schwartzentruber DJ, Weber JS, Ettinghausen SE, White DE, Steinberg SM, Cole DJ, Kim HI *et al.*: **Randomized comparison of high-dose and low-dose intravenous interleukin-2 for the therapy of metastatic renal cell carcinoma: an interim report.** *J Clin Oncol* 1994, **12**:1572-1576.
127. Klapper JA, Downey SG, Smith FO, Yang JC, Hughes MS, Kammula US, Sherry RM, Royal RE, Steinberg SM, Rosenberg S: **High-dose interleukin-2 for the treatment of metastatic renal cell carcinoma: a retrospective analysis of response and survival in patients treated in the surgery branch at the National Cancer Institute between 1986 and 2006.** *Cancer* 2008, **113**:293-301.
128. Rosenberg SA, Lotze MT, Muul LM, Leitman S, Chang AE, Ettinghausen SE, Matory YL, Skibber JM, Shiloni E, Vetto JT *et al.*: **Observations on the systemic administration of autologous lymphokine-activated killer cells and recombinant interleukin-2 to patients with metastatic cancer.** *N Engl J Med* 1985, **313**:1485-1492.
129. Ahmadzadeh M, Rosenberg SA: **IL-2 administration increases CD4+ CD25(hi) Foxp3+ regulatory T cells in cancer patients.** *Blood* 2006, **107**:2409-2414.
130. Rosenberg SA, Yang JC, Sherry RM, Kammula US, Hughes MS, Phan GQ, Citrin DE, Restifo NP, Robbins PF, Wunderlich JR *et al.*: **Durable complete responses in heavily pretreated patients with metastatic melanoma using T-cell transfer immunotherapy.** *Clin Cancer Res* 2011, **17**:4550-4557.
131. Rosenberg SA, Packard BS, Aebersold PM, Solomon D, Topalian SL, Toy ST, Simon P, Lotze MT, Yang JC, Seipp CA *et al.*: **Use of tumor-infiltrating lymphocytes and interleukin-2 in the immunotherapy of patients with metastatic melanoma. A preliminary report.** *N Engl J Med* 1988, **319**:1676-1680.
132. Dudley ME, Yang JC, Sherry R, Hughes MS, Royal R, Kammula U, Robbins PF, Huang J, Citrin DE, Leitman SF *et al.*: **Adoptive cell therapy for patients with metastatic melanoma: evaluation of intensive myeloablative chemoradiation preparative regimens.** *J Clin Oncol* 2008, **26**:5233-5239.